Broad spectrum biocontrol of plant pathogens with Bacillus amyloliquefaciens strains through selective enhancement of its antibiotic production using carbon sources and culture additives

Saghar Mazarei^{1,2}, L. Chien^{1,2}, C.Y. Yuan^{1,2}, K. Dumont^{2,4}, S. Tagliabracci⁵, S. Kandasamy², R. Nicol⁵, S. Saldias², G.

Lazarovits^{1,2}

¹Department of Biology, Western University, London, ON, ²A&L Biologicals, Agroecological Research Services Centre, London, ON ³Department of Biotechnology, Canadore College, North Bay, ON, ⁴ Research & Innovation, Lambton College, Sarnia ON



Introduction

- Fungal pathogens are the major cause of pathogen-related yield loss in agriculture, threatening global food security¹
- Although fungicides are used to mitigate such plant diseases, chemical fungicide use has become less effective due to the emergence of fungicide resistance in pathogen populations²
- Thus, utilization of bacterial species as sustainable and natural alternatives continues to receive considerable attention
- Bacillus amyloliquefaciens are promising as potential biocontrol species, as nearly 8% of their genome is involved in antimicrobial metabolite synthesis³
- Interestingly, Bacilli show differential lipopeptide production depending on culture conditions⁴

Objectives –

- Increase the metabolite synthesis of three Bacillus strains (94, 113, 279) through the addition of various culture additives such as growth hormones and carbon sources
- Evaluate the biocontrol ability of the amended cultures against economically relevant plant pathogens

1- PCR Amplification of antibiotic ketides 2- Creation of 110 amended cultures with various carbon sources and organic acids 3- antagonistic assay against 19 fungal isolates 4- LC-MS Differential Profiling of Antibiotics 5- Greenhouse and field trials against

pathogens of interest

Results

In vitro Antagonistic Assay

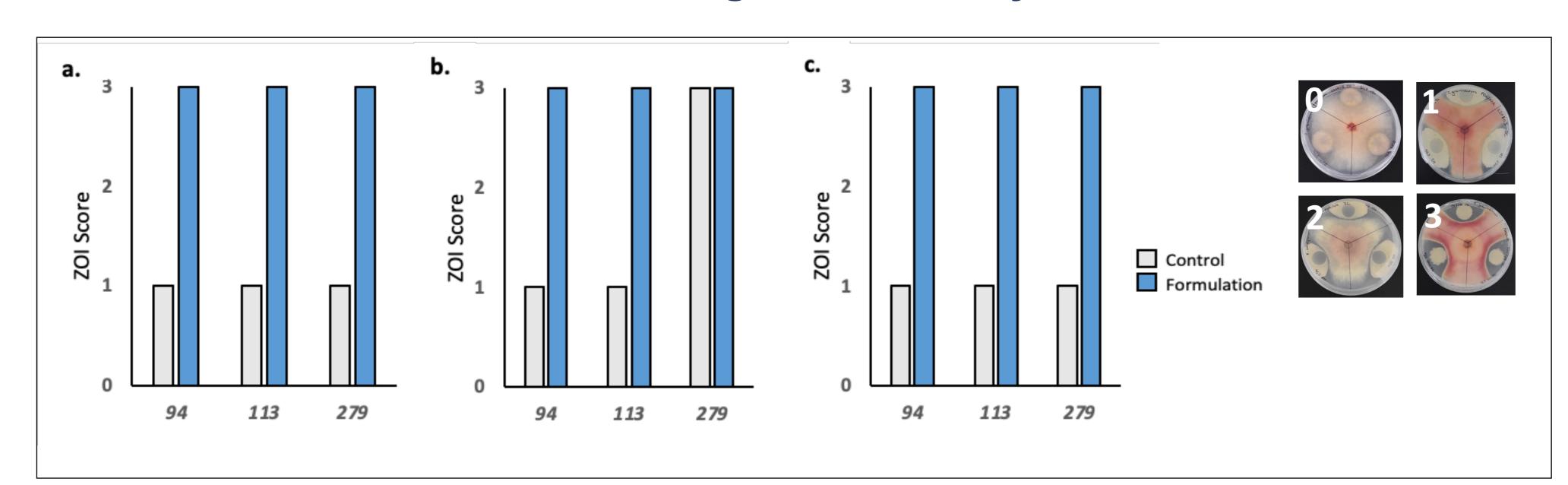


Fig. 1. in vitro antagonistic abilities of control and amended isolates against Fusarium oxysporum (a), Fusarium graminearum (b), and Slerotinia sclerotiorum (c). The isolates were assigned a score between 0 to 3 based on their zone of inhibition (ZOI), as seen in the example (d).

LC-MS Differential Screening

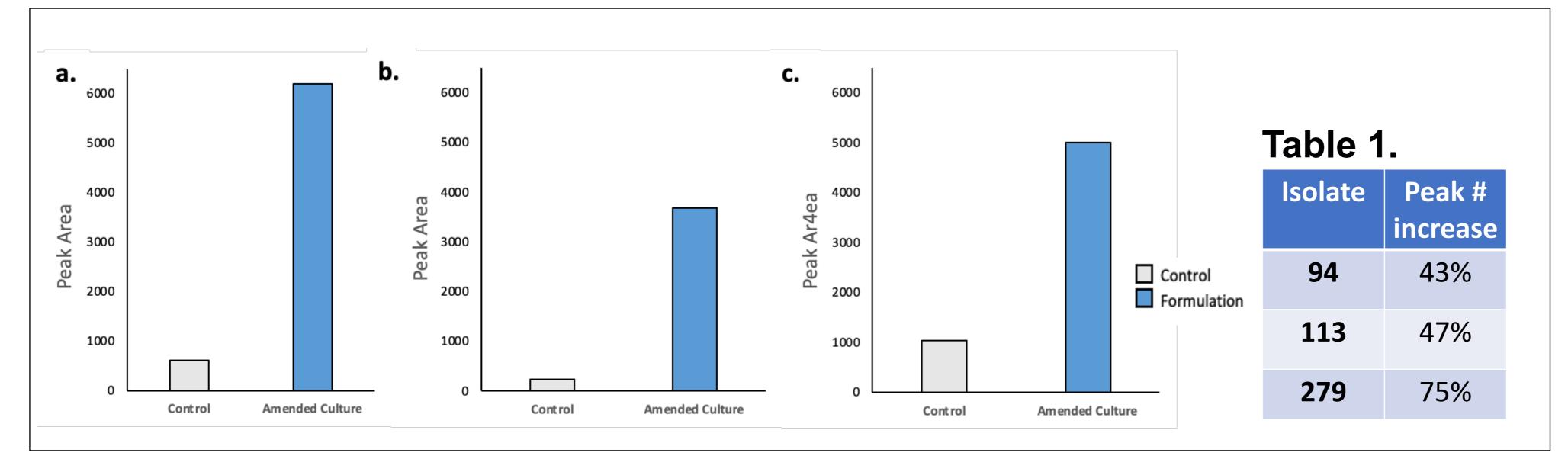


Fig. 2. LC-MS results for amended cultures and respective controls. The total peak area for 94 (a), 113 (b), and 279 (c) are on the left, and the total peak number increase for the three isolates is exhibited in **table 1**.

Conclusion

- All three *Bacillus amyloliquefaciens* strains showed improved lipoprotein production upon manipulation of growth media⁴; confirmed with LC-MS data (**Fig. 2.**)
- Amended cultures with highest lipoprotein production demonstrated increased antagonism against pathogens, notably *F. oxysporum*, *F. graminearum*, and *S. sclerotiorum* (**Fig. 1**)
- Modified cultures improved plant survival (**Fig. 4)** and delayed symptom expression (**Fig. 5**)
- Thus, the strains could potentially be used as natural biocontrol alternatives, mitigating issues associated with fungicide resistance²

Greenhouse & Field Trial

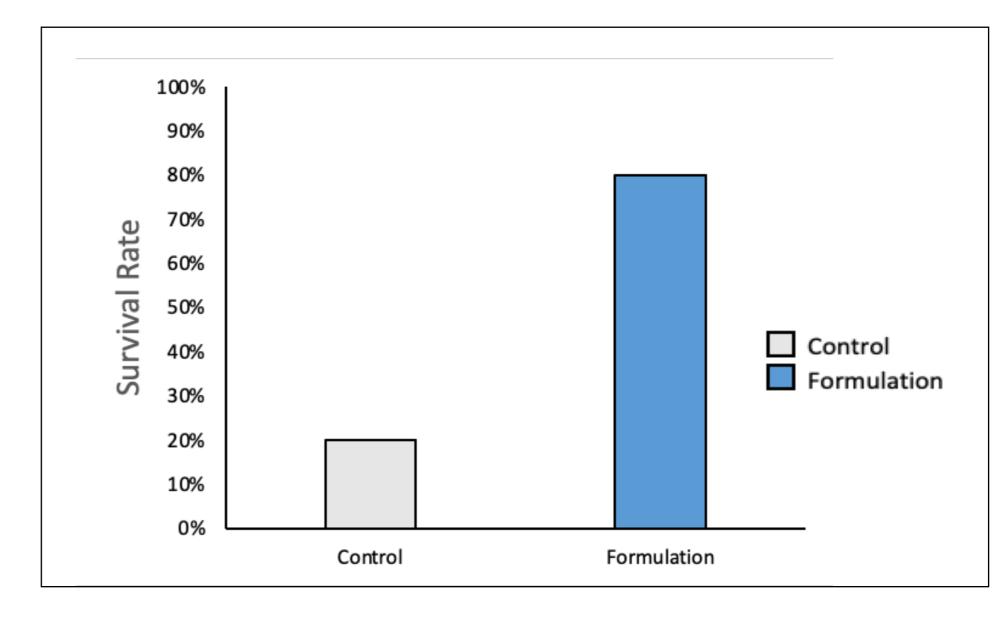


Fig. 4. Growth room trial data for tomatoes artificially infected with *Fusarium oxysporum*. The plants were either treated with amended 94, or left untreated as control; plant survival rate was quantified after 14 days.

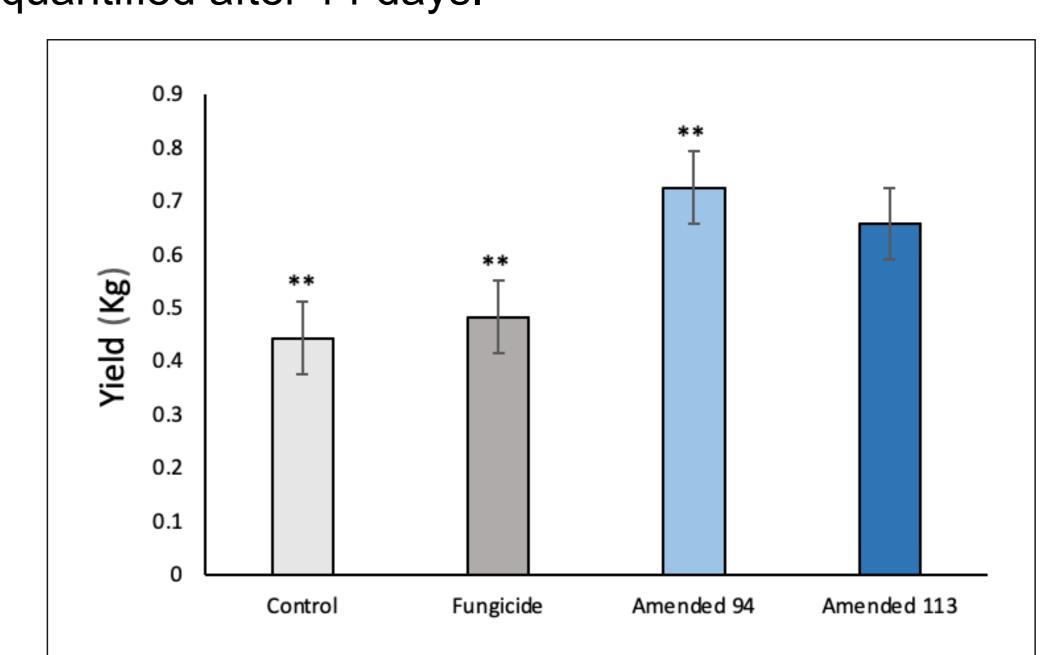


Fig. 5. Average yield of field squash infected with powdery mildew. Plants were treated with either amended cultures or a common chemical fungicide. An equal number were left untreated as control.

Future Directions –

- Organize further growth room trials against additional pathogens of interest
- Conduct assays to quantify pathogen load throughout greenhouse and field trials; collect quantitative data
- Repeat field experiments to confirm preliminary results

References

Brazilian Journal of Microbiology, 39(2), 286–295. https://doi.org/10.1590/s1517-83822008000200017

¹⁾ Fisher, M. C., Henk, D. A., Briggs, C. J., Brownstein, J. S., Madoff, L. C., McCraw, S. L., & Gurr, S. J. (2012) Emerging fungal threats to animal, plant and Ecosystem Health. *Nature*, 484(7393), 186–194.

https://doi.org/10.1038/nature10947

2) Deising, H. B., Reimann, S., & Pascholati, S. F. (2008). Mechanisms and significance of fungicide resistance.

³⁾ Chen, X.-H., Koumoutsi, A., Scholz, R., & Borriss, R. (2008). More than anticipated – production of antibiotics and other secondary metabolites by *bacillus amyloliquefaciens* FZB42. *Microbial Physiology*, *16*(1-2), 14–24. https://doi.org/10.1159/000142891

^{4).} Akpa, E., Jacques, P., Wathelet, B., Paquot, M., Fuchs, R., Budzikiewicz, H., & Thonart, P. (2001). Influence of culture conditions on lipopeptide production by bacillus subtilis. *Applied Biochemistry and Biotechnology*, 91-93(1-9), 551–562. https://doi.org/10.1385/abab:91-93:1-9:551